OFGS File: P/717-181(CONT)

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Patent Application of:

Alessandra BOE, et al.

Date: October 13, 2000 Serial No.: Unknown

Filed: Herewith

For: TNF RECEPTOR AND STEROID HORMONE IN A COMBINED THERAPY

Asst. Commissioner of Patents and Trademarks Washington, DC 20231

REQUEST FOR CONTINUING APPLICATION UNDER 37 C.F.R. 1.53(b)

Sir:

This is a request for the filing of a Continuation application under the provisions of 37 C.F.R. 1.53(b) of pending application Serial No. 08/983,223, filed February 27, 1998, by Alessandra Boe and Francesco Borrelli, entitled "TNF RECEPTOR AND STEROID HORMONE IN A COMBINED THERAPY". The prior application is hereby incorporated by reference.

Enclosed is a copy of the prior application, including the oath or declaration as originally filed.

I hereby state that the attached papers are a copy of prior application Serial No. 08/983,223, filed February 27, 1998, without any new matter therein.

Cancel in this application original claims 2-17 of the prior application before calculating the filing fee.

The filing fee is calculated as follows:

BASIC Filing Fee: \$ 710.00

Number of Claims in Excess of 20: _0 x \$18

Number of Independent Claims over 3: 0 x \$80

One or more multiple dependent claims: \$260

TOTAL FILING FEE: <u>\$ 710.00</u>



Check No. 001654 which includes the amount of \$710.00 in payment of the filing fee is enclosed herewith.

The Patent and Trademark Office is hereby authorized to charge any additional fees or credit any refund, at any time during the prosecution of this application, to Deposit Account No. 15-0700.

Amend the specification by inserting at page 1, line 3, the paragraph:

--This is a continuation of application Serial No.

08/983,223, filed February 27, 1998.--

This application includes THREE (3) sheets of drawings containing Figures 1 through 2b.

Priority under 35 U.S.C. § 119 based on PCT Application No. PCT/EP95/02767, filed July 14, 1995, is claimed. The certified copy of the priority application has been filed in said prior application Serial No. 08/983,223, filed February 27, 1998.

The prior application was assigned to Applied Research Systems ARS Holding N.V. and is recorded at Reel 9623, Frame 0117.

The power of attorney in the prior application, as originally filed, is to customer no. 2352, OSTROLENK, FABER, GERB & SOFFEN, LLP, 1180 Avenue of the Americas, New York, New York 10036-8403, and the members of the firm: Reg. No. 17,542; Samuel H. Weiner, Reg. No. 18,510; Jerome M. Berliner, Reg. No. 18,653; Robert C. Faber, Reg. No. 24,322; Edward A. Meilman, Reg. No. 24,735; Stanley H. Lieberstein, Reg. No. 22,400; Steven I. Weisburd, Reg. No. 27,409; Max Moskowitz, Reg. No. 30,576; Stephen A. Soffen, Reg. No. 31,063; James A. Finder, Reg. No. 30,173; William O. Gray, III, Reg. No. 30,944; Louis C. Dujmich, Reg. No. 30,625, and Douglas A. Miro, Reg. No. 31,643, as attorneys with full power of substitution and revocation to prosecute this application, to transact all business in the Patent and Trademark Office in connection therewith and to receive all correspondence. The Power appears in the original papers in the prior application.

00475185.1

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Dorothy Jenkins

Name of Person Mailing Correspondence

October 13, 2000

Date of Signature

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IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re: Patent Application of

Alessandra BOE, et al. Date: October 13, 2000

Serial No.: Unknown Group Art Unit: --

Filed: Herewith Examiner: --

For: TNF RECEPTOR AND STEROID HORMONE IN A COMBINED THERAPY

Asst. Commissioner for Patents Washington, D.C. 20231

PRELIMINARY AMENDMENT

Sir:

00474729.1

Preliminary to examination, please amend the above-identified application as follows:

IN THE SPECIFICATION:

Please amend the specification as follows:

Page 3, line 7, change "Applicant has" to --Applicants have--.

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Page 4, line 13, after "therapy" insert --,--.

Page 5, line 20, after "specified" insert --,--.

Page 6, line 6, after "by" insert --the--.

Page 7, line 5, after "μ/mouse)" insert --,--; line 32, after group" insert --,--.

Page 8, line 13, after "experiments" insert --,--.

Page 9, line 6, after "allows" insert --one--.

IN THE ABSTRACT

Please amend the abstract as follows:

Line 8, delete the paragraph indication.

IN THE CLAIMS:

Plesae cancel claim 1 without prejudice and substitute therefor the following new claims:

- --18. A method for treating lethal viral infections in a patient which comprises administering to a patient a tumour necrosis factor (TNF) receptor in combination with dehydroepiandrosterone (DHEA) simultaneously, separately or sequentially.
 - 19. The method of claim 18, wherein the TNF receptor is TNF Binding Protein-2.
 - 20. The method of claim 18, wherein the TNF receptor is TNF Binding Protein-1.
- 21. A method for treating autoimmune and inflammatory diseases in a patient which comprises administering to the patient a tumour necrosis factor (TNF) receptor in combination with dehydroepiandrosterone (DHEA) simultaneously, separately or sequentially.
 - 22. The method of claim 21, wherein the TNF receptor is TNF Binding Protein-2.

00474729.1

- 23. The method of claim 22, wherein the disease treated is autoimmune disease.
- 24. The method of claim 22, wherein the disease treated is inflammatory disease.
- 25. The method of claim 21, wherein the TNF receptor is TNF Binding Protein-1.
- 26. The method of claim 25, wherein the disease treated is autoimmune disease.
- 27. The method of claim 25, wherein the disease treated is inflammatory disease.
- 28. The method of claim 21, wherein the disease treated is autoimmune disease.
- 29. The method of claim 21, wherein the disease treated is inflammatory disease.--

REMARKS

The claims in this application have been amended to appropriate condition for consideration and allowance.

The early examination and allowance of this application is respectfully solicited.

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October 13, 2000

Date of Signature

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Respectfully submitted,

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TNF RECEPTOR AND STEROID HORMONE IN A COMBINED THERAPY

FIELD OF THE INVENTION

The present invention relates to the use of a TNF Receptor together with a steroid hormone to produce a pharmaceutical composition for the treatment of lethal bacterial and viral infections as well as autoimmune and inflammatory diseases. It also relates to said pharmaceutical compositions for the simultaneous, separate or sequential use of its active ingredients for the above specified treatment.

In particular, it relates to the use of TBP-1 together with dehydroepiandrosterone (DHEA) or its metabolites to produce a pharmaceutical composition for the treatment of septic shock.

BACKGROUND OF THE INVENTION

Septic shock as a consequence of Gram-negative bacteremia or endotoxemia remains a critical clinical condition in spite of adequate antibiotic therapy.

It is now known that the lethal consequences of septic shock results from an exaggerated host response, mediated by protein factors such as TNF and interleukin 1 (IL-1), rather than from the pathogen directly.

Tumour necrosis factor (TNF) is a cytokine produced mainly by activated macrophages, which elicits a wide range of biological effects. These include an important role in endotoxic shock and in inflammatory, immunoregulatory, proliferative, cytotoxic and anti-viral activities.

The induction of the various cellular responses mediated by TNF is initiated by its interaction with two distinct cell surface receptors of approximately 55 kDa (also called TNF-R1) and 75 kDa (also called TNF-R2). The extracellular soluble portions of these receptors, called respectively TBP-1 (TNF Binding Protein-1) and TBP-2 (TNF Binding Protein-2), have been isolated and cloned (see EP Patents 308 378, 398 327 and EP patent Application 433 900).

Several studies in animal models of TNF-mediated endotoxic shock indicated that both anti-TNF antibodies and soluble TNF-receptor are able to counteract the lethal effects induced (see for example: Bentler, B et al., Science,

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229:869 (1985), Lesslauer, W. et al., Eur. J. Immunol., 21:2883 (1991), Evans, T.J. et al. J. Exp. Med., 180:2173 (1994) and Mohler K.M. et al., J. Immunol. 151:1548 (1993)).

The *in vivo* protective effects of urinary as well as recombinant TBP-1 (derived from both CHO and *E. Coli* cells) in experimental models of septic shock have already been demonstrated (see Bertini, R. et al., Eur. Cytokine Netw. 4(1):39 (1993) and Ythier A., et al., Cytokines, 5:459 (1993)).

DHEA (INN: Prasterone) is an adrenocortical steroid hormone which is an intermediate in the biosynthesis of other hormones including testosterone and estradiol-17B.

The precise biological functions of DHEA are still unclear. Experimental and epidemiological data suggest an inverse relationship between low levels of DHEA in serum and morbidity from atherosclerotic cardiovascular disease (see Barret-Connor, D. et al., N. Engl. J. Med. 315:1519 (1986)), cancer (see Gordon, G.B., et al., Cancer Res. 51:1366 (1991)) and immunodeficiency virus (HIV) infection (Villette, J.M. et al. J. Clin, Endocrinol, Metab. 70:572 (1990)).

An immunomodulating activity of this drug has also been reported; in particular. DHEA has been shown to prevent the development of systemic lupus erythematosus in a mouse model (Lucas, J. et al. J. Clin. Invest. 75:2091 (1985)).

It has been demonstrated that DHEA regulates the systemic resistance against lethal viral infections induced by different viruses: coxsackievirus B4 (as reported in Loria, R.M. et al., Ann. N.Y. Acad. Sci. 293), herpes virus type 2 (see Loria, R.M. et al., J.Med. Virol., 26:301 (1988)), West Nile virus (a neurovirulent Sindbis virus) and Semliki Forest virus (as reported in Ben-Nathan, D. et al. Arch. Virol. 20:263 (1989)).

It has also been reported that DHEA has similar protective effects against a lethal bacterial infection induced by *Enterococcus faecalis* (see Loria, R.M. et al. in Symposium Pharmaco-Clinique, Roussel-Uclaf 9:24 (1989)).

Danenberg, H.D. et at. (Antimicrob. Agents Chemother.36:2275 (1992)) reported that DHEA has the capability of protecting mice from septic shock induced by lipopolysaccharides (LPS) alone or by Tumor Necrosis Factor alpha (TNF- α) in combination with D-Galactosamine. LPS administration resulted in high levels of TNF- α , a response that was significantly blocked by DHEA, both in vivo and in vitro.

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DISCLOSURE OF THE INVENTION

The main object of the present invention is the use of a TNF Receptor in combination with a steroid hormone to produce a pharmaceutical composition for the treatment of lethal bacterial and viral infections as well as autoimmune and inflammatory diseases. The TNF Receptor and the steroid hormone can be administered simultaneously, separately or sequentially.

The Applicant has, in fact, found that in such treatment there is a synergic effect between the two active ingredients.

Another object of the present invention is, therefore, the method for treating lethal bacterial and viral infections as well as autoimmune and inflammatory diseases by administering simultaneously, separately or sequentially an effective amount of TNF Receptor and an effective amount of a steroid hormone, together with a pharmaceutically acceptable excipient.

A further object of the present invention are the pharmaceutical compositions containing a TNF Receptor and a steroid hormone, in the presence of one or more pharmaceutically acceptable excipients, for the simultaneous, separate or sequential administration of its active ingredients in the treatment of lethal bacterial and viral infections as well as autoimmune and inflammatory diseases.

In case of separate or sequential use of the two active ingredients, the pharmaceutical compositions of the invention will consist of two different formulations, each comprising one of the two active ingredients together with one or more pharmaceutically acceptable excipients.

The administration of such active ingredients may be by intravenous, intramuscular or subcutaneous route. Other routes of administration, which may establish the desired blood levels of the respective ingredients, are comprised by the present invention.

Non-limitative examples of pathologies, in which the above active ingredients are indicated, are the following: septic shock, AIDS, rheumatoid arthritis. lupus erythematosus and multiple sclerosis. Particularly preferred is the treatment of septic shock.

The TNF Receptor is preferably selected between the extracellular soluble domain of TNF-R1, i.e. TBP-1, and the extracellular soluble domain of TNF-R2, i.e. TBP-2. TBP-1 is particularly preferred. Recombinant human TBP-1 (r-hTBP-1) is advantageously used according to the invention.

The steroid hormone can be both a corticosteroid or an androgen. Preferably, it is an androgen. More preferably, it is DHEA or one of its metabolites, dispersed in a suitable medium, for example a phospholipid emulsion or carboxymethyl cellulose or polyvinylpirrolidone.

Therefore, a preferred embodiment of the invention consists in the combined use of r-hTBP-1 and DHEA in the treatment of septic shock. In this case, the Applicant has found that it is possible to reduce at least four times the effective dose of r-hTBP-1.

The above effect has been showed with in vivo experiments in mice.

In particular, a murine model has been used in the present invention, according to which septic shock is induced by administering lipopolysaccharides (LPS) in combination with D-Galactosamine.

For the human therapy the preferred doses of the active ingredient are 20 mg of r-hTBP-1 and 80 mg of DHEA, preferably divided into four administrations in a 24-hours period.

The invention will now be described by means of the following Examples, which should not be construed as in any way limiting the present invention. The Examples will refer to the Figures specified here below.

20 BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1: Protective effects of CHO r-hTBP-1 (62.5 μ g/mouse) in combination with single (250 μ g/mouse) or repeated injections (4 x 62.5 μ g/mouse) of DHEA in the experimental murine septic shock are shown.

<u>Figure 2</u>: Protective effects of different doses of CHO r-hTBP-1 in the experimental murine septic shock induced by LPS + D-Galactosamine are shown for comparative purposes. The number of mice in the r-hTBP-1 groups is the same as that reported for the corresponding control groups. In Figure 2a and 2b are shown the results observed after 24 and 48 hours (respectively) after the inoculum of LPS + D-Galactosamine.

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EXAMPLE 1

LPS + D-Galactosamine-induced septic shock model

The experiments were carried out *in vivo* on mice. The septic shock was induced by an i.p. administration of a mixture of LPS $(0.1 \mu g/mouse)$ and D-Galactosamine (18 mg/mouse).

The treatment schedule for r-hTBP-1 based on repeated administrations (the total dose was divided into 4 administrations given at time 0 and 3. 6 and 24 hours after), was adopted. CHO r-hTBP-1 was tested, alone or in combination with DHEA, at dose levels of 62.5 and 125 µg/mouse (total dose).

5 Assessment of the effects of DHEA in the LPS + D-Galactosamine model

DHEA was tested both in a single administration injected i.p. one hour before and contemporaneously to the septic shock-inducing agent at doses encompassed between 100 and 2.000 µg/mouse and in repeated treatment with the total dose (250 µg/mouse) divided into 4 injections given within 24 hours.

10 Assessment of the effects of DHEA in combination with r-hTBP-1

The above treatment schedules of DHEA (doses from 250 to 1,500 μ g/mouse) were followed also in combination with r-hTBP-1 which was tested at the total dose varying from 62.5 to 125 μ g/mouse (divided into 4 administrations).

15 Materials

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<u>Animals</u>

SPF female BALB/c inbred mice, 6-8 weeks (about 18-20 g b.w.), from Charles River were used throughout the whole study after an acclimatisation period of at least 7 days.

Unless otherwise specified in all the experiments the mice were randomly divided into groups of 10 animals each.

Test drugs

CHO recombinant human TBP-1 produced according to the known methods in the field of biotechnology.

The protein mass of the r-hTBP-1 bulk was determined by the Lowry method before starting the experiments.

E. Coli lipopolysaccharides 055:B5 supplied by Sigma.

D-(+)-Galactosamine hydrochloride, 99% supplied by Janssen.

Dehydroepiandrosterone supplied by Sigma.

30 Reagents

Sodium chloride 0.9% (Saline), manufactured by Baxter.

Phospholipid emulsion (PLE) composed of phosphatidyl choline (71%), phosphatidyl ethanolamine (16%), lysophosphatidyl choline (7.5%), sphingomyelin (5%) and phosphatidic acid (0.3%).

Methods

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Septic shock model (LPS + D-Galactosamine model)

Both substances were dissolved in saline. The mixture of LPS (0.1 μ g/mouse) and D-Galactosamine (18 mg/mouse) was prepared by mixing equal volumes of these substances. The mixture was subsequently injected to animals by i.p. route. This dose was selected under previous experiments as that inducing around 80% mortality calculated as the number of dead vs. total number of mice for each group. The lethality was monitored after 24, 48, 72 and 96 hours and at day 7 post-treatment.

10 Treatment schedule for r-hTBP-1

The drug was diluted in saline and injected by repeated administrations, i.e. i.v. immediately before the inoculum of LPS plus D-Galactosamine and s.c. 3, 6 and 24 hours later.

Treatment schedules for DHEA

The effects of DHEA, alone or in combination with r-hTBP-1, were assayed after single i.p. administrations at different times in respect to LPS + D-Galactosamine injection; in particular, it was administered 1 hour before and contemporaneously to the septic shock-inducing agent.

A repeated treatment schedule was also investigated in which DHEA was administered at time 0 (septic shock induction) and 3, 6 and 24 hours later. DHEA was initially vehicled in a mixture of 2.5% alcohol and 10% rabbit serum in saline. To improve the solubility and standardise the preparation, the steroid was successively dispersed in a mixture of 2.5% alcohol and 10% phospholipid emulsion in saline.

25 Data evaluation

The primary end point was the survival rate at 48 hours after induction of septic shock. The comparison of the effects of each treatment group vs. controls and among treatment groups was effected by Fisher's exact test, one-tailed. The same comparison was also performed at the other time points. The results in each treatment group were expressed as percent survival.

Results

DHEA administered 1 hour before LPS + D-Galactosamine

The results of the DHEA dose-range finding experiments are reported in Table 1. No protective effects were exerted by this substance up to 1,000 μ g/mouse; by contrast, statistically significant protective effects were obtained at

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higher doses with a complete protection at 2.000 µg/mouse at all time points considered.

When a suboptimal dose of CHO r-hTBP-1 (125 µg/mouse), inducing about 50% survival, was administered in combination with two suboptimal doses of DHEA (1.000 and 1.500 µg/mouse) an additive effect between the two substances was observed (Table 2). The result of the combination with the highest DHEA dose, where a 100% survival rate was obtained, is significantly different from that of the r-hTBP-1 group (50%) and DHEA group (40%).

DHEA administered contemporaneously with LPS + D-Galactosamine

The results obtained in the DHEA dose-range finding test, where the substance, diluted in saline 10% rabbit serum, was administered contemporaneously to septic shock induction are reported in Table 3.

Although a clear dose-effect relationship has not been found, a higher protection was produced by 1.000 µg DHEA in comparison to what observed with the same dose by the previous treatment schedule (DHEA administered 1 hour before).

The combination of 62.5 µg/mouse of r-hTBP-1 with 250 µg/mouse of DHEA induced a slightly higher protection (60%) in respect to the single treatments (20% and 30% for r-hTBP-1 and DHEA, respectively, see Table 4). When the same combination was vehicled in a 10% phospholipid emulsion, to improve the solubility of DHEA, a significantly higher survival was obtained compared to DHEA and r-hTBP-1 alone (Table 4) at 24 hours which, however, successively decreased.

Repeated administrations of DHEA after LPS + D-Galactosamine

Comparable results (30 and 40% survival rate at 48 hours) were obtained when a dose of 250 µg/mouse of DHEA was administered as a single injection (Table 5, group D) or divided into 4 mocula (given at 0, 3, 6 and 24 hours from septic shock induction, group E), respectively. Similarly, no statistically significant differences were found when the animals were injected the combination of r-hTBP-1 and DHEA with the latter administered either as a single inoculum (Table 5, group F) or divided into 4 injections (Group G).

Due to the high survival rate obtained in the r-hTBP-1 group an additive effect is not evident in these tests after 24 and 48 hours where the combination-treated groups were significantly different only from control and DHEA groups.

However, it has to be stressed that the combined therapy induced a more

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prolonged survival in respect to the r-hTBP-1 group alone since a statistically significant difference was found after 72 and 96 hours.

Figure 1 reports the data obtained combining the results of the three experiments reported in Tables 4 and 5 in which the dose of 250 µg/mouse of DHEA was injected as a single inoculum or divided into 4 inocula (62.5 µg each).

Figure 2 reports, for comparative purposes, the effects of different doses of CHO r-hTBP-1 alone in the experimental murine septic shock induced by LPS + D-Galactosamine. The number of mice in the r-hTBP-1 groups is the same as that reported for the corresponding control groups. In Figure 2a and 2b are shown the results observed after 24 and 48 hours (respectively) after the inoculum of LPS + D-Galactosamine.

In all the experiments percent survival values were calculated using the survival Tables for the *follow up* studies (Armitage P., Cap. XIV, Tavole di sopravvivenza in Statistica Medica. Feltrinelli, Milano). The χ^2 -test was applied to each observation time by comparing the DHEA + r-hTBP-1 groups to the other groups.

In addition to showing that at all time points the combined therapy is significantly different from control and DHEA groups, this analysis confirmed that more prolonged survival times were obtained with the combined treatment in respect to r-hTBP-1 alone. Although no statistical difference between the two treatment combinations has been found, a trend to a more sustained protection can be observed when both drugs were divided into four injections, which is reflected by the different levels of significance at different time-points (Table 6).

25 Conclusions

A significant protection from death (70% survival) was found with at least 1.500 µg/mouse of DHEA, injected 1 hour before LPS + D-Galactosamine, which confirm literature data.

When suboptimal doses of DHEA (1,000 and 1,500 µg/mouse) and of r-hTBP-1 (125 µg/mouse, total dose) were combined, additive effects (90-100% survival) were produced, compared to 0-40% and 50% with the two doses of DHEA and of r-hTBP-1, respectively.

Similar results were obtained even with lower doses of DHEA (250 μ g/mouse) in combination with r-hTBP-1 (62.5 μ g/mouse) (90-100% survival), which also provided a more prolonged protection (up to 96 hours) in respect to r-

hTBP-1 alone, both when DHEA was injected contemporaneously to LPS + D-Galactosamine in a single inoculum (250 μ g/mouse) or when the same dose was divided into 4 inocula (62.5 μ g/each) injected at times 0, 3, 6 and 24 hours after septic shock induction.

The present findings are of particular relevance in showing that a combined treatment of r-hTBP-1 with DHEA allows to reduce the dose of r-hTBP-1 by at least four times.

EXAMPLE 2

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Example of pharmaceutical manufacturing

A) r-h-TBP-1

15 Materials

Pure saccharose Ph EurBP. Ph Nord. NF. (Merck); H₃PO₄ Suprapur (Merck); NaOH for analysis (Merck); water for injectable.

Vials DIN 2R (borosilicate glass type I), rubber closures (Pharmagummi W1816 V50) and Aluminium rings and Flip-off caps (Pharma-Metal GmbH) are used as containers.

<u>Preparation of r-hTBP-1 solution containing saccharose</u> (for 1,000 vials each containing 5 mg of r-hTBP-1)

Saccharose (30.0 g) and H_3PO_4 (1.96 g) are dissolved into water for injectables (800 ml) in order to obtain the starting saccharose solution.

The bulk of r-hTBP-1 (5 g) is added to the saccharose solution that, after the pH has been adjusted at 7.0 by means of 2.5 M NaOH, is brought to the final volume of 1.000 ml. The solution is filtered through a 0.22 µm Duarapore sterile filter (Millipore). During the process, the solution temperature is kept between 4° and 8°C.

Filling up and lyophilisation

The vials are filled up with 1 ml of r-hTBP-1 sterile solution, transferred to the freeze dryer and cooled at -45°c for 6 hours. The lyophilisation is started at the temperature of -45°C with a vacuum of 0.07 mBar. The heating is performed

according to the following scheme: 10°C for 12 hours; then +35°C until the end of the cycle.

B) DHEA

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Materials

Pure Mannitol Ph Eur. BP. FU. USP. FCC. E241 (Merck): H₃PO₄ Suprapur (Merck): NaOH for analysis (Merck): water for injectable.

Vials DIN 2R (borosilicate glass type I), rubber closures (Pharmagummi W1816 V50) and Aluminium rings and Flip-off caps (Pharma-Metal GmbH) are used as containers.

Preparation of DHEA solution containing mannitol (for 1.000 vials containing 10 mg of DHEA)

Mannitol (45.0 g) and H_3PO_4 (1.96 g) are dissolved into water for injectables (800 ml) in order to obtain the starting saccharose solution.

The bulk of DHEA (20 g) is added to the saccharose solution that, after the σH has been adjusted at 7.0 by means of 2.5 M NaOH, is brought to the final volume of 1.000 ml. The solution is filtered through a 0.22 μm Durapore sterile filter (Millipore). During the process, the solution temperature is kept between 4° and $8^{\circ}C$.

Filling up and lyophilisation

The vials are filled up with 1 ml of DHEA sterile solution, transferred to the freeze dryer and cooled at -45°c for 6 hours. The lyophilisation is started at the temperature of -45°C with a vacuum of 0.07 mBar. The heating is performed according to the following scheme: +20°C for 12 hours: then +35°C until the end of the cycle.

Table 1 - Effects of DHEA in the septic shock induced by LFS + D-Galactosamine, in mice

1		No.				% cumulat	% cumulative survival				
Treat	Treatment,	of	24	24 hrs	48	18 hrs					- 1
dose / mou.	dose / mouse and route	mico	!					72 hrs	96	96 hrs	
		82	181	2nd	1st	2nd	151	2nd	151	2nd	
			cxh	дхө	exp	бхр	exo	2.0			
	None	10	20	20	20	01	300	450	dxa	фхө	
LPS 0.1 µg,	DHEA, i p.	10	20			2	07	0	. 20	10	
i.p.(1)	100 µg (2)				77		50	,	20		
+	DHEA in							į		-	
	7	2	2	1	10		10				-
	500 µg (2)		X	-			2		2		
D-Gal. 18 mg,	DHEA, i.p.	10		30							
i.p.(1)	1 000 нд. (2)			0.7	t	70	,	10	ı	0	
	DHEA in	10									
	1 500 µg. (2)	2	1	p 0/	ı	70 a	ı	90	4	909	
	. 42170										
	UHEA, I.p.	01	100 a	100 a	100a	100 a	1003				1
	2 000 µg. (2))	3	001	100 a	100 a	100 a	

(1)Administered as a mixture at time 0;

(2)Administered 1 hour before LPS + D-Gal administration

(a)Significantly different from LPS + D-Gal controls (Fisher's exact test, one-tailed) N.B. DHEA was vehicled in saline 10% rabbit serum

Table 2 - Effects of CHO r-hTBP-1 and DHEA in septic shock induced by LPS + D-Galactosamine, in mice

					•	
Treatm	Treatment, dose/mouse and route	NO 04				
		5		% cumular	% cumulative survival	
		Mice	24 hrs			
	None	10	Z. III.S	48 hrs	72 hrs	96 hrs
		2	10	10	10	
	r-hТВР-1 125 µg (2)	10	50	~ · ·	01	10
LPS, i.p.	DHEA 1 000 :: : 131		00	90	20	50
0.1 (1)	is di la	0	0	0		
0.1 Jug. 11.					>	0
	DHEA 1 500 no in (3)					
+		≘	40	40	20	06
	The same the same and the same and the same same same same same same same sam					07
	DHEA 1 000 µg, i.p. (3)				*****	
D-Gal, i.p.	+	10	ao a b			
18 mg, (1)	r-hTBP-1 125 µg (2)	-		90 a, n	90 a, b	90 a, b
	DHEA 1 500 µg, i.p. (3)					
	†	10	100 a, b, c	100 a b c		
	r-hTBP-1 125 μg (2)				100 ª, u, c	100 a, b, c

(2) Administered at time 0 (i.v.), and at 3, 6 and 24 hours (s.c.) after LPS + D-Gal injection. (1) Administered as a mixture at time 0;

(3) Administered 1 hour before L.PS + D-Gal administration;

(b) Significantly different from DHEA alone; (c) Significantly different from r-(a) Significantly different from LPS + D-Gal. controls;

hTBP-1 alone (Fisher's exact test, one-tailed)

N.B.- DHEA was vehicled in saline 10% rabbit serum.

Effects of DHEA in the septic shock induced by LPS and D-Galactosamine, in mice Table 3 -

Treat	Treatment,	No			%	% cumulative survival	ive surviv	ļe,		
esnow/esop	and route	of mice	24	24 hrs	48	48 hrs	72	72 hrs	96	96 hrs
			1st	2nd	1st	2nd	1st	2nd	151	2nd
			e (p.	ехр.	өхр.	өхр.	вхр.	вкр.	екр.	exp.
	None	10	10	40	10	40	10	40	10	40
1000										
LPS 0.1 µg,	DI LEA, i.p.	10	•	20	,	20	,	20		
i.p.(1)	250 µg (2)					ì		0.7	•	70
	DHEA, i.p.	10	20	30	40	30	06			
T	500 µg. (2)				2	3	OS S	00	90	9
D-Gal. 18 mg,	DHEA, i.p.	10	40	50	40	50	70			
i.p.(1)	750 µg (2)				}	3	P F	000	4 0	90
	DHEA, i.p.	10	80a		- 09		GO		00	
	1 000 µg. (2)				1		3	,	00	,

(1) Administered as a mixture at time 0

(2) Administered at time 0

a Significantly different from LPS + D-Gal controls (Fisher's exact test, one-tailed)

N.B.: DHEA was vehicled in saline 10% rabbit serum

Table 4 - Effects of CHO r-hTBP-1 and DHEA in the septic shock induced by LPS and D-Galactosamine, in mice

	Treatment,	No.	% (umulativ	e surviv	9/
dose / mou	se and route	of	24	48	72	96
		mice	hr	hr	hr	hr
	none	20	25	25	25	25
	r-hTBP-1, 62.5 μg (2)	10	20	10	0	0
LPS 0.1µg	DHEA 250 μg (3)	10	30	30	30	30
ı.p. (1)	NaCl 10%RS)					
-	DHEA 250 μg (3)					
D-Gal 18mg	(NaCl 10%RS)	10	60	50	50	50
i.p. (1)	-			İ	į	
	r-hTBP-1 62.5 µg (2)					
	DHEA 250 μg (3)	10	40	30	30	30
	(NaCl 10% PLE)					
	DHEA 250 μg ⁽³⁾					
	(NaCl 10% PLE)	10	90 a,b,c	60 c	40 C	40 °
	-					
	r-hTBP-1 62.5 µg (2)					

⁽¹⁾ Administered as a mixture at time 0

⁽²⁾ Administered at time 0 (i.v.) and at 3, 6 and 24 hours (s.c.) after LPS + D-Gal. injection.

⁽³⁾ Administered as a single injection at time 0 i.p.

⁽a) Significantly different from LPS + D-Gal. controls (Fisher's exact test, one-tailed).

⁽b) Significantly different from DHEA alone (Fisher's exact test, one-tailed).

⁽c) Significantly different from r-hTBP-1 alone (Fisher's exact test, one-tailed).

N.B. DHEA was vehicled in saline 10% rabbit serum (NaCl 10% RS) or in saline 10% phospholipid emulsion (NaCl 10% PLE).

Table 5 - Effects of CHO r-hTBP-1 and DHEA in the septic shock induced by LPS + Galactosamine, in mice

	Tr.	Treatment	No.			6	-frianto				
	dose/mo	dose/mouse and route	,				canna	A Cumulative Stirvival	al		
			à	24	24 hrs 	48	48 hrs	72	72 hrs	96	96 hrs
			mice	151	2nd	751	2nd	151	2nd	151	2,00
				exo.	өхр	67.0	2				7117
	<	anon	,			ides	cxp.	вхр.	ехр.	өхр.	exp.
301			2	2	10	10	10	10	1	1	5
5	20	NaCi 10% PLE	10	,	-				2	2	2
)		=	1	=	·	=	ı	=
					(n = 9)		(n - 0)				
0.1 μα,	ن	r.hTBP-1 62 E (2)	,				16-111		(n - 9)		(n = 9)
		12 61 6.20	10	90a	70a	80a	40	203	5		
.p.	۵	DHEA 250 Hg (3)	101	•			2	-0/	ا ا	709	30
-	L		2	0+	•	30		30	'	30	
_	ונ	UHEA 62.5µg x4 (4)	10	40	9	()				2	.
D-Gal.	Ľ	DHEA 250 (3)		2	04	40	40	40	40	40	40
		יייבר בטס ווק יכי									
la mg,		+	10	100a,b	ı	100a.b		4			
i.p. (1)		r-hTBP-1 62.5 µg (2)					,	1004,0		100a,b	,
	<u>.</u>	PUEA CO F. 141									
	,	DITEM 02.5119 x4 191			-						
		+	10	100a,b	90a,b	100a.b	heoo	4		•	
		r-hTBP-1 62.5 ug (2)					2000	100a,0	80a,c	100a,b	80a'c
T.1.		6				-					

The notes to the table are reported in the next page

Notes to Table 5

- (1) Administered as a mixture at time 0
- (2) Administered at time 0 i.v., and at time 3, 6 and 24 hours (s.c.)
- (3) Administered as a single injection at time 0 i.p.
- (4) Administered at time 0, 3, 6 and 24 hours i.p.
- (a) Significantly different from LPS + D-Gal. controls (Fisher's exact test, one-tailed).
- (b) Significantly different from DHEA alone (Fisher's exact test, one-tailed).
- (c) Significantly different from r-hTBP-1 alone (Fisher's exact test, one-tailed).
- N.B. DHEA was vehicled in 10% phospholipid emulsion (NaCl 10% PLE)

Table 6 - Statistical significance of r-hTBP-1 group vs. r-hTBP-1 + DHEA groups (χ²-test)

Observation times	p l	'evel
(hours post LPS + D- Gal injection)	r-hTBP-1 vs. r-hTBP-1 62.5 + DHEA 250 (1)	r-hTBP-1 vs. r-hTBP-1 62.5 + DHEA 4x62.5 (1)
24	0.015	0.015
48	0.022	0.0006
72	0.024	0.0002
96	0.024	0.0002

(1) Doses are expressed as µg/mouse.

CLAIMS

- Use of a TNF Receptor in combination with a steroid hormone to produce a
 pharmaceutical composition for the treatment of lethal bacterial and viral
 infections.
- 2. Use of a TNF Receptor in combination with a steroid hormone to produce a pharmaceutical composition for the treatment of autoimmune and inflammatory diseases.
- 3. The use according to Claim 1 or 2, wherein the TNF Receptor and the steroid hormone are used simultaneously, separately or sequentially.
- 4. The use according to any preceding claim, wherein the TNF Receptor is selected between the extracellular soluble domain of TNF-R1, TBP-1 and the extracellular soluble domain of TNF-R2, TBP-2.
- 5. The use according to any preceding claim, wherein the TNF Receptor is the extracellular soluble domain of TNF-R1. TBP-1.
- 6. The use according to any preceding claim, wherein the steroid hormone is selected between a corticosteroid and an androgen.
- 7. The use according to claim 6, wherein the steroid hormone is an androgen.
- 8. The use according to claims 6 or 7, wherein the androgen is DHEA.
- 9. The use according to claim 1, wherein the lethal bacterial infection is septic shock.
- 10. Pharmaceutical composition containing a TNF receptor and a steroid hormone, in the presence of one or more pharmaceutically acceptable excipients, for the simultaneous, separate or sequential use of its active ingredients in the treatment of lethal bacterial and viral infections.
- 11. Pharmaceutical composition containing a TNF receptor and a steroid hormone, in the presence of one or more pharmaceutically acceptable excipients, for the simultaneous, separate or sequential use of its active ingredients in the treatment of autoimmune and inflammatory diseases.
- 12. The pharmaceutical composition according to claim 10 or 11, wherein the TNF Receptor is selected between the extracellular soluble domain of TNF-R1, TBP-1 and the extracellular soluble domain of TNF-R2, TBP-2.
- 13. The pharmaceutical composition according to claims 10 or 12, wherein the TNF Receptor is the extracellular soluble domain of TNF-R1, TBP-1.

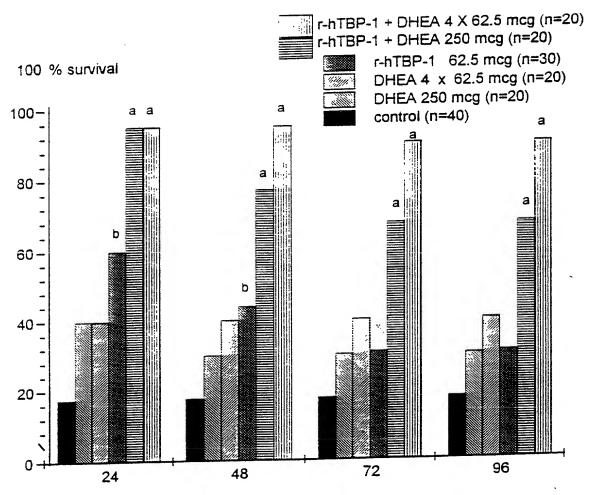
- 14. The pharmaceutical composition according to any of claims 10 to 13, wherein the steroid hormone is selected between a corticosteroid and an androgen.
- 15. The pharmaceutical composition according to any of claims 10 to 14, wherein the steroid hormone is an androgen.
- 16. The pharmaceutical composition according to any of claims 10 to 15, wherein the androgen is DHEA.
- 17. The pharmaceutical composition according to claim 10, wherein the lethal baterial infection is septic shock.

ABSTRACT

The present invention relates to the use of a TNF Receptor together with a steroid hormone to produce a pharmaceutical composition for the treatment of lethal bacterial and viral infections as well as autoimmune and inflammatory diseases. It also relates to said pharmaceutical compositions for the simultaneous, separate or sequential use of its active ingredients for the above specified treatment.

In particular, it relates to the use of TBP-1 together with dehydroepiandrosterone (DHEA) or its metabolites to produce a pharmaceutical composition for the treatment of septic shock.

Figure 1

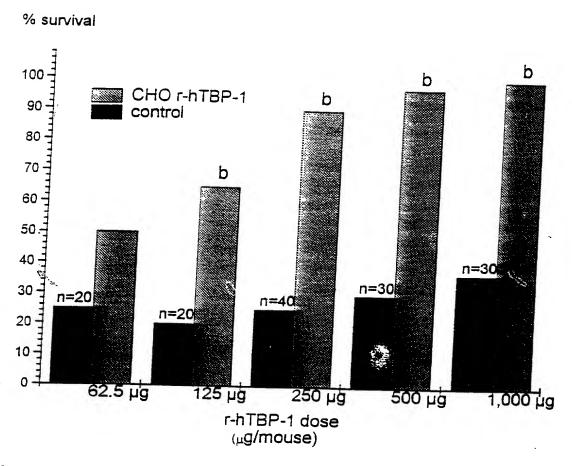


hours post LPS + D-Galactosamine injection

a = significantly different from r-hTBP-1 group (χ^2 -test) b = significantly different from control group (χ^2 -test)

Figure 2 a

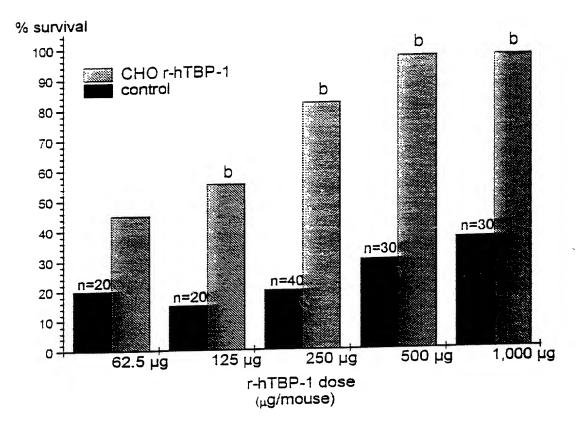
24 hours post LPS + D-Gal injection



 $b = significantly different from control group (<math>\chi^2$ -test)

Figure 2 b

48 hours post LPS + D-Gal injection



 $b = significantly different from control group (<math>\chi^2$ -test)

COMBINED DECLARATION	UNITED STATES		PATENT	`APPLICA	TION		s file no. 7-/54
As a below named inventor, I I that I verily believe that I am the o named) of the subject matter which TNF RECEPTOR AN	nginal, first and sole i is claimed and for w	inventor (it only on which a patent is sou	e name is ght on th	s listed belo e invention	w) or a joir entitled:	as stated below at inventor (if plu	next to my name; ural inventors are
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the specification of which is attached was filed on 14 Jul application number PC?	d hereto, unless the ly 1995	following box is che as Ui	cked: uited State	es patent A _I	pplication N	umber or PCT I	nternational patent
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I hereby state that I have review any amendment referred to above. I acknowledge the duty to disch Regulations, §1.56. I hereby claim priority benefits United States provisional application having a filing date before that of the	wed and understand the ose all information known and under Title 35, Unite (s) listed below and the application on whi	nown to be material ed States Code §119 have also identified	to patent	tified specif ability in ac	ication, incl cordance w	uding the claims ith Title 37, Cod	i, as amended by
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I hereby claim the benefit under subject matter of each of the claims paragraph of Title 35, United States Title 37. Code of Federal Regulatio international filing date of this appli	of this application is Code, §112, I acknowns, §1.56 which beca						
UNITED STATES APPLICATION NUMBER		DATE OF FILING				STATI	US .
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I hereby appoint customer no. 2: Reg. No. 18,510; Jerome M. Berlin Stanley H. Lieberstein - Reg. No. 2: Reg. No. 31,063; James A. Finder- Douglas A. Miro - Reg. No. 31,643 business in the Patent & Trademark	352 OSTROLENK, F er - Reg. No. 18,653 2,400; Steven I. Weis Reg. No. 30,173; W , as attorneys with fi Office connected the	FABER, GERB & S; Robert C. Faber - sburd - Reg. No. 27 Villiam O. Gray, III all power of substitute with and to receive	OFFEN, Reg. No ,409; Ma - Reg. N tion and	LLP, and to 24,322; Ex Moskowi (o. 30,944; revocation tespondence	he members dward A. N tz - Reg. N Louis C. D o prosecute	s of the firm, San Meilman - Reg. I o. 30,576; Steph ujmich - Reg. N this application,	nuel H. Weiner - No. 24,735; ten A. Soffen - o. 30,625 and to transact all
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